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**A-140 ATTWOOD'S METHOD FOR AMNIOTIC
FLUID EMBOLISM**

FIXATION: Formalin, 10% Buffered Neutral (F-113)

SECTION: Paraffin @ 6 microns

STAINING PROCEDURE: (Use control slide)

1. Deparaffinize and hydrate to distilled water.
2. **Alcian Green Solution, 1 %**(A-140-1), for two minutes. Rinse in distilled water.
3. **Potassium Permanganate, 0.5% Aqueous** (A-140-3), for two minutes. Rinse in distilled water.
4. **Oxalic Acid, 5%, Aqueous** (A-140-4) for two minutes. Rinse in distilled water.
5. **Mayer's Hematoxylin** (A-140-5), for 3 minutes.
6. Rinse and blue in **Scott's Water Solution** (A-140-6), for 5 minutes.
7. **Phloxine-Calcium Chloride Solution** (A-140-2), for 30 minutes. Rise in distilled water, then in Cellosolve.
8. Differentiate each slide with **Tartrazine Cellosolve Solution** (A-140-9), checking until the Phloxine is removed from Erythrocytes.
9. Dehydrate in Cellosolve, clear in **Xylene** (C-120), two changes each.
10. Mount with **Permount** (M-18).

RESULTS:

Nuclei.....blue-black
Mucin, Ground Cartilage Substance,
Granules of certain Mast Cells.....green
Squamous Epithelium, Fibrin.....red
Erythrocytes, Plasma, Collegen.....yellow
Inclusion Bodies.....red

REFERENCES:

AFIP Manual of Histological Staining Methods, 3rd. ed., Ed. L. Luna: New York: McGaw Hill Publications, c. 1968, p. 232.

Attwood, H.D., J. Path. Bact., 76:211-215, 1958