

E-306 LAQUEUR'S METHOD FOR ALCOHOLIC HYALIN

Fixation: 10% Buffered Neutral Formalin (F-113) or Zenker's

Sections: Paraffin at 6 microns

Staining:

1. Deparaffinize and hydrate to distilled water.
2. Stain in **Mayer's Hematoxylin** (E-306-1) for 15 minutes. Wash in running water for 15 minutes.
3. Flood slides with **Acid Fuchsin - Aniline Solution** (E-306-2), *shake bottle well before using solution will separate upon standing*, and heat gently until aniline fumes escapes; *view against a dark background*, Let stand for 5 minutes. Wash in water, 3 dips.
4. Differentiate in **Alcoholic Picric Acid** (E-306-5) until only hyalin and red cells remain red, and collagen is faint gray or unstained. Wash in running water for 3 minutes.
5. Mordant in 1% Phosphomolybdic Acid (E-306-4) for 4 to 18 hours.
6. Counterstain in Light Green Solution (E-306-3) for 1 hour. Wash in running water for 1 minute.
7. fibers are seen as distinct fibrils.
8. Dehydrate in 95% alcohol, absolute alcohol, and clear in Xylene, two changes each.
9. Mount with **Permount** (M-18).

Results:

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| Mallory Bodies | Bright red |
| Erythrocytes | Red |
| Cytoplasm | Pale brown |
| Bile pigment | Green |
| Proteinaceous material seen occasionally in some liver cells | Red |
| Hemosiderin and lipofichsin | Unstained |

References:

Laqueur, G.L., *Amer. J. Clin. Path.*, 20:689-690, 1950.

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Lillie, R.D., Fullmer, H.M., *Histopathologic Technic and Practical Histochemistry* 4th ed., McGraw-Hill, N.Y., 1976, pp670-671.

Clark, G., (ed.) *Staining Procedures*, 3rd edition, Williams & Wilkins, Baltimore, p. 52 c. 1973